resection of the tumor is a typical method of treatment.
ADVANTAGE - The microspheres can pass through a narrow-gauge needle and can be delivered to tumor beds without systemic side-effects.
pp; 22 DwgNo 1/17
Derwent Class: A96; B07; D16; P32; P73
International Patent Class (Main): A61K-009/50
International Patent Class (Additional): A61F-002/02; B01J-013/02; B32B-005/16

012933745
WPI Acc No: 2000-105592/200009
Enteric coated pharmaceutical composition for arresting the release of the drug from orally ingestible dosage forms
Patent Assignee: BRISTOL-MYERS SQUIBB CO (BRIM)
Inventor: ULLAH I; WILEY G J
Number of Countries: 080 Number of Patents: 003
Patent Family:

Patent No	Kind	Date	Applicat No	Kind	Date	Week	
WO 9961002	A1	19991202	WO 98US16128	A	19980804	200009	B
AU 9886854	A	19991213	AU 9886854	A	19980804	200020	
BR 9815861	A	20010116	BR 9815861	A	19980804	200107	
			WO 98US16128	A	19980804		

Priority Applications (No Type Date): US 9883597 A 19980522
Patent Details:

Patent No	Kind	Lan	Pg	Main IPC	Filing Notes
WO 9961002	A1	E	30	A61K-009/16	
AU 9886854	A			A61K-009/16	Based on patent WO 9961002
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Designated States (National): AL AM AT AU AZ BA BB BG BR BY CA CH CN CU CZ DE DK EE ES FI GB GE GH GM HU ID IL IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MD MG MK MN MW MX NO NZ PL PT RO RU SD SE SG SI SK SL TJ TM TR TT UA UG UZ VN YU ZW
Designated States (Regional): AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW NL OA PT SD SE SZ UG ZW

Abstract (Basic): WO 9961002 A1
NOVELTY - The high drug load composition includes a medicament which may degrade in a low pH environment but which is protected from enteric coating. The composition comprises a core in the form of beadlet, pellet, granule or particle and an enteric coating for the core.
DETAILED DESCRIPTION - **An enteric coating composition comprises a core in the form of a beadlet, granule or particle and an enteric coating for the core.** The core comprises 50-100 weight % (wt.%) acid labile medicament, 0-10 wt.% binder and 0-10 wt.% disintegrant. **The enteric coating further comprises a methacrylic acid copolymer and a plasticizer.** The coating imparts protection to the core so that the core is afforded the protection in a low pH environment of 3 or less while capable of releasing medicament at a pH of 4.5 or higher. The composition also comprises 0.1-4 wt.% anti-adherent. An INDEPENDENT CLAIM is included for a process of preparing an enteric-coated composition comprises: (a) preparing a dry blend containing a medicament, a binder and a disintegrant, and setting a portion of the dry blend aside; (b) forming a wet mass from the remainder of the dry

blend not set aside in (a); (c) extruding the wet mass to form an extrudate and spheronizing the extrudate into high-potency beadlets by dusting the wet mass extrudate with the portion of the dry blend set aside in (a); (d) coating the beadlets with an enteric coating polymer and plasticizer in an aqueous media; and (e) blending the coated beadlets with an anti-adherent.

USE - The invention is used for arresting the release of the drug from orally ingestible dosage forms.

ADVANTAGE - The invention provides excellent protection in very acidic environment (pH less than 3) while not delaying the rapid release in regions of pH greater than 4, whether this be the upper intestine or the duodenum. The process not only eliminates the costly additional subcoating step, but also allows quicker release of the drug since the added subcoat layer delays drug release. The process allows very high drug loads and would not change the composition of the bead, regardless of the amount of dry blend used for dusting. It is also involves the preparation of a dry blend of powdered drug substance with or without a very small amount of suitable binder and optional disintegrant. The enteric coating employed is easier to process, and is especially advantageous for coating small diameter, low mass particles (beadlets) with minimal processing problems (agglomeration) without the need for organic solvents.

pp; 30 DwgNo 0/1

Derwent Class: A14; A96; B07; D16

International Patent Class (Main): A61K-009/16

International Patent Class (Additional): A61K-009/54; A61K-009/62

012506509

WPI Acc No: 1999-312614/199926

Enteric coated granules for lactic acid bacteria

Patent Assignee: IL YANG PHARM CO LTD (ILYA-N); IL YANG PHARM IND CO LTD (ILYA-N)

Inventor: JEON H R; KIM D Y; PARK D W

Number of Countries: 024 Number of Patents: 003

Patent Family:

Patent No	Kind	Date	Applicat No	Kind	Date	Week
WO 9920745	A1	19990429	WO 98KR314	A	19981016	199926 B
KR 99032308	A	19990515	KR 9753312	A	19971017	200030
EP 1023440	A1	20000802	EP 98947986	A	19981016	200038
			WO 98KR314	A	19981016	

Priority Applications (No Type Date): KR 9753312 A 19971017

Patent Details:

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WO 9920745	A1	E	25	C12N-011/02	

Designated States (National): CA CN JP US

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KR 99032308	A		A23C-009/13	
EP 1023440	A1	E	C12N-011/02	Based on patent WO 9920745

Designated States (Regional): AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE

Abstract (Basic): WO 9920745 A1

NOVELTY - Enteric coated granules for lactic acid bacteria comprise a lactic acid bacterial seed, a water-miscible coating and an optional

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(74) Agents: CHOI, Kyu, Pal et al.; 824–20, Yeoksam–dong, Kangnam–ku, Seoul 135–080 (KR).

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(54) Title: ENTERIC COATED MICROGRANULES FOR STABILIZING LACTIC ACID BACTERIA

(57) Abstract

The present invention relates to an enteric coated granule prepared by coating lactic acid bacteria–containing seed with a water–miscible coating material and then, if desired, subjecting the first coated product to the second coating with a controlled–release coating material.

ENTERIC COATED MICROGRANULES FOR STABILIZING LACTIC ACID BACTERIA

TECHNICAL FIELD

The present invention relates to an enteric coated microgranule for optimally stabilizing lactic acid bacteria. In the present specification, the term "lactic acid bacteria" means the bacteria beneficial to health, which are present in human intestine and help to keep the peristalsis of intestine active.

BACKGROUND ART

The ingested lactic acid bacteria prevent the abnormal fermentation of food and activate the function of intestine, thereby improving the functional abnormality of intestine such as constipation, diarrhea, etc. and maintaining good care of health. Also, the addition of lactic acid bacteria to feedstuff can prevent the accumulation of gas, constipation, diarrhea, etc. caused by the abnormal fermentation inside the intestine of livestock which may result from the repetitive supply of the same feedstuff, which ultimately improves the quality of flesh and highly contributes to the development of dairy farming.

However, in spite of high usefulness and values of lactic acid bacteria, the actual use of lactic acid bacteria has many restrictions due to their acid-unstability. That is, since lactic acid bacteria are very unstable under pH 4, almost all the ingested lactic acid bacteria are destructed at the acidity of gastric juice (about pH 2). Therefore, only a trace amount of the ingested lactic acid bacteria (about one per million) can reach the intestine alive. As a result, much time and

expenses are required to make lactic acid bacteria efficiently exhibit their functions in human intestine.

In order to overcome such a problem, a way to increase the amount of lactic acid bacteria which reach the intestine by using more than 10 times excess of bacteria has been proposed in the field of food and pharmaceutical industry. However, it is not a fundamental solution, but merely a very fragmentary and wasting, temporary remedy. Further, food containing microcapsules wherein lactic acid bacteria are mixed with fat, emulsifying agent and protective material and then encapsulated has recently been reported, the purpose of which is to increase the ratio of lactic acid bacteria arrived at the intestine by making the bacteria survive in gastric juice (see, Korean Patent Laid-open Publication No. 97-25405). However, according to the experimental result, it has been identified that upon ingestion of such encapsulated lactic acid bacteria their coating is disintegrated within 30 minutes regardless of the circumstance being gastric juice or intestinal juice. It can also be noted from other experiments that the commercially available lactic acid bacteria coated with gelatin as the coating base are not disintegrated for more than 10 hours in any circumstances without difference on the specificity to gastric juice or intestinal juice (see, Experimental Example 1). It appears that this is because the material used as the coating base for lactic acid bacteria is a conventional one which does not react sensitively to the property of the gastric or intestinal juice. Further, numerous organic solvent-based coating methods utilizing various polymers have been reported in the general pharmaceutical field (see, PCT/JP94/001675, Japanese Patent Appln. Nos. 91-235667, 92-364123, 92-41434, 93-186335, 93-186336, etc.). However, such coating techniques are not satisfactory to protect lactic acid bacteria from gastric juice. Particularly, if organic solvent is used as a solvent of the coated preparation or if the coating

process is carried out at a high temperature of more than 55℃, the actual survival rate of lactic acid bacteria in human body is much less than the expected value.

On the other hand, it has been tried to develop variant strains of lactic acid bacteria, which have a high acid-resistance. However, this approach requires greater time and cost, and has worse effect than the coating methods.

DISCLOSURE OF INVENTION

The present inventors have intensively studied about enteric coating technique which gives some usable merits in view of the stability of lactic acid bacteria and in the economical view. As a result, we have found that when lactic acid bacteria are first coated with a specific water-miscible coating material and then, if desired, second coated with a conventional controlled-release coating material, the destruction of lactic acid bacteria during the procedure for preparing the coated granule can be greatly reduced and, furthermore, the coated granule capable of delivering lactic acid bacteria contained therein to the target organ in which lactic acid bacteria actually display their function, i.e. intestine, by safely protecting lactic acid bacteria from the attack of gastric juice can be produced. Thus, we have completed the present invention. In the present invention, since the coated granule contains active lactic acid bacteria in a high ratio and is very sensitive to acidity, the bacteria contained therein can survive under human gastric circumstance and the granule then can be disintegrated rapidly in the intestine.

Therefore, it is an object of the present invention to provide an enteric coated microgranule specially designed so as to display the

function of lactic acid bacteria in the intestine by optimally stabilizing lactic acid bacteria contained in the granule.

BEST MODE FOR CARRYING OUT THE INVENTION

The coated granule containing lactic acid bacteria according to the present invention is more specifically explained in below.

The coated granule containing lactic acid bacteria according to the present invention can be prepared by first coating the lactic acid bacteria-containing seed with a water-miscible coating material at low temperature and, if desired, then subjecting the first coated product to the second coating with a controlled-release coating material. In the present invention, the destruction of lactic acid bacteria during the procedure for preparation can be minimized by conducting the first coating with a water-miscible material at low temperature.

In the present invention, one or more strains beneficial to human being, which are selected from the group consisting of *Streptococcus* genus, *Lactococcus* genus, *Leuconostoc* genus, *Pediococcus* genus, *Enterococcus* genus, *Lactobacillus* genus and *Bifidobacterium* genus can be used as the lactic acid bacteria strain.

The water-miscible coating material which can be used for the first coating includes sodium alginate as the main ingredient of seaweed (e.g., brown seaweed) extract, alginic acid, polymethylmethacrylate [Eudragit L30D, Eudragit LS30D, Kollicoat MAE 3DP (manufactured by BASF Co.), etc.], wheat protein, soybean protein, methylcellulose (MC), hydroxypropylcellulose (HPC), hydroxypropylmethylcellulose [HPMC; pharma coat, aqua coat, etc.], polyvinylacetatephthalate [Sureteric;

manufactured by Colorcon Co.], gums, for example, guar gum, locust bean gum, xanthan gum, gellan gum, arabic gum, etc. Since these water-miscible coating materials are water-soluble or water- dispersible, it is advantageous that the first coating procedure can be conveniently carried out by using water as a solvent. This is very important in view of the fact that any organic solvent which is not only harmful to human body but also fatal to the stability of lactic acid bacteria is not used for the coating procedure, and therefore, the problem of removing the residual organic solvent is completely solved. This is a characteristic advantage of the present invention as distinct from the prior techniques which necessarily require the use of organic solvents to dissolve high-molecular substances as a coating material. In the present invention, sodium alginate is preferably used as the water- miscible first coating material. The reason is that since sodium alginate is water-soluble and its aqueous solution is neutral, it is much more advantageous for the stability of lactic acid bacteria.

The seed used for the coating procedure can be either lactic acid bacteria themselves or a mixture of lactic acid bacteria and one or more additive substances selected from the group consisting of starch, lactose, oligosaccharides, glycoalcohols, calcium gluconate, calcium lactate and gluconic acid. These additives are added for the purpose of diluting lactic acid bacteria in a desired ratio, activating only lactic acid bacteria while suppressing other bacteria strains, or improving the proliferation of lactic acid bacteria.

In the first coating procedure, the water-miscible coating material is preferably used in an amount of 1 to 80% by weight with respect to the seed.

Although the first coated granule of lactic acid bacteria as prepared above is sufficiently effective by itself, but it can be more effectively used after second coating with a conventional controlled-release coating material. Therefore, an enteric coated microgranule with both the first and second coatings is also included within the scope of the present invention.

As the second coating material, the controlled-release coating material, particularly an enteric coating material commonly used in pharmaceutical field; or a coating material for swelling such as carbopol or arabic gum; and other controlled-release coating materials can be used. More specifically, corn protein extract (described in USP/NF) and artificial processed materials thereof, such as for example, Zein-DP or prolamin, sodium alginate, alginic acid, polymethylmethacrylate, for example, Eudragit L30D, Eudragit LS30D, Kollicoat MAE 3DP (manufactured by BASF Co.), etc., shellac, hydroxypropylmethylcellulose phthalate (HPMCP), hydroxypropylmethylcellulose (HPMC), hydroxy-propylmethylcelluloseacetatesuccinate (HPMCAS), carboxymethylcellulose (CMC), hydroxypropylcellulose (HPC), celluloseacetatephthalate (CAP), polyvinylacetatephthalate [Sureteric(Colorcon Co.)], ethylcellulose (EC), methylcellulose (MC), soybean protein or wheat protein (they are registered as Food Additives), chitin, chitinic acid, agar, carrageenan, pectin, carbopol, or gums, such as for example, guar gum, locust bean gum, xanthan gum, gellan gum, arabic gum, etc. can be mentioned. Among them, one or more selected from the group consisting of corn protein extract, hydroxypropylmethylcellulose phthalate (HPMCP) and shellac are preferably used as the second coating material.

In conducting the second coating procedure, one or more materials selected from the coating materials as mentioned above are used in an

amount of 1 to 95% by weight with respect to the first coated granule. Particularly, when the enteric coating material commonly used in the pharmaceutical field is used, it is used in an amount ranging from 1 to 40% by weight; or when other coating materials for swelling are used, it is used in the amount ranging from 30 to 95% by weight. The kind and amount of the coating material may be appropriately determined by a person skilled in the relevant technical field, considering the property of coating material and the purpose of using the coating material.

Contrary to the first coating wherein only water is used as a solvent for the protection of lactic acid bacteria, the second coating may use one or more various solvents selected from water, alcohol, acetone, acetonitrile, methylene chloride, ether, hexane, chloroform, 1,4-dioxane, tetrahydrofuran, dimethylsulfoxide, ethyl acetate and methyl acetate. In case the coating material is hardly dissolved in the solvent, if required, a pH regulator such as acetic acid, hydrochloric acid, phosphoric acid, various buffer solutions, citric acid, tartaric acid, malic acid, etc. can be used to adjust the pH to the desired range thereby improving the solubility of the coating material. This can be easily carried out by a person skilled in the relevant art.

When both of the first and second coatings are conducted, the coating materials used in the respective steps should be different from each other. If desired, one or more plasticizers selected from a group consisting of polyethyleneglycols, myvacet, propyleneglycol, glycerine, triethyl citrate, triacetin, cetyl alcohol and stearyl alcohol can be used in the first or second coating procedure. In this case, the plasticizer is preferably used in an amount of 1 to 50% by weight with respect to the coating material as used.

To optimally stabilize the lactic acid bacteria in preparing the

coated granule, the present invenors have utilized a process in which (a) the seed containing lactic acid bacteria is suspended and, at the same time, spray-coated with a coating-solution, or (b) the seed suspended in the coating-solution is dispersed into a chamber. Thus, the present invention can be carried out more preferably by applying such processes.

The coating process may be carried out by using a fluidized bed granulator, CF-granulator and the like, preferably a fluidized bed granulator (SFC-MINI, Freund co., Japan). When such a granulator is used, the temperature of the introduced air is preferably maintained in the range of 40 to 70℃. The temperature of granule in the granulator at each step should be kept more than 20℃ to prevent the granule from absorbing moisture from the ambient atmosphere and coagulating with each other. Preferably, the temperature of granule is maintained from 25 to 55℃ throughout the whole procedures since lactic acid bacteria may be destroyed at the temperature exceeding 55℃.

The present invention will be more specifically explained by the following examples and experimental examples. However, it should be understood that the examples are intended to illustrate but not to in any manner limit the scope of the present invention.

Example 1

(A) First Coating

Seed : *Lactobacillus acidophilus*:
Lactobacillus bifidus:
Streptococcus faecalis
= 1:1:1(w/w/w) mixture 250g

Coating-Solution :	Sodium alginate	3g
	Water	300㎖

(B) Second Coating

Seed : Coated granule according to (A)		253g
Coating-Solution:	Zein-DP(processed from corn protein extract)	50g
	Cetanol	5g
	80% Ethanol	500㎖
	Glycerine	5㎖

i) Preparation of first coated granule

Lactic acid bacteria were suspended in a fluidized bed granulator (SFC-MINI, Freund co., Japan) and, at the same time, spray-coated with the first coating-solution as given above. The operation conditions of the granulator were adjusted to the values given in the following Table 1. Particularly, the temperature of lactic acid bacteria-containing powder, that is the coated powder, in the granulator was carefully controlled not to deviate from the temperature ranging from 25 to 55℃.

ii) Preparation of second coated granule

The first coated granule according to the above procedure i) was suspended in a fluidized bed granulator (SFC-MINI, Freund co., Japan) and, at the same time, spray-coated with the second coating-solution consisting of Zein-DP and 80% ethanol. Glycerine was further added to the coating-solution as a plasticizer. The operating conditions of the granulator were adjusted to the values given in the following Table 1.

Particularly, the temperature of lactic acid bacteria-containing powder, that is the coated powder, in the granulator was carefully controlled not to deviate from the temperature ranging from 25 to 55℃.

Table 1.

	First Coating	Second Coating
Temp. of Introduced Air(℃)	60	60
Temp. of Granule in granulator(℃)	30	35
Flow rate of Introduced Air(㎥/min)	9	9
Flow rate of Excreted Air(㎥/min)	10	10
Flow rate of Introduced Air / Slit (㎥/min)	7	7
Flow rate of Introduced Air / Fluid (㎥/min)	7	7
Spray rate of Coating-Solution (㎖/min)	10	12
Flow rate of Sprayed Air(㎥/min)	35	35
Rotation Number of Rotor(rpm)	300	300
Rotation Number of Agitator(rpm)	500	500
Rotation Number of Lump Breaker (rpm)	2500	1700
Spur Jet (on-off)	20 sec. each	20 sec. each

Example 2

The coated granule according to the present invention was prepared according to the same procedure as Example 1 except that materials as described below were used.

(A) First Coating

Seed :	*Lactobacillus acidophilus*: *Lactobacillus bifidus*: *Streptococcus faecalis* = 1:1:1(w/w/w) mixture	25g
	Lactose	225g
Coating-Solution :	Sodium Alginate	3g
	Water	300㎖

(B) Second Coating

Seed :	Coated granule according to (A)	253g
Coating-Solution:	Liquid Shellac (Opaglos®; Colorcon Co.)	30㎖
	Zein-DP(processed from corn protein extract)	25g
	Glycerine	5.0㎖
	80% Ethanol	300㎖

Example 3

The coated granule according to the present invention was prepared according to the same procedure as Example 1 except that materials as described below were used.

(A) First Coating

Seed : *Lactobacillus acidophilus*: *Lactobacillus bifidus*: *Streptococcus faecalis* = 1:1:1(w/w/w) mixture		250g
Coating-Solution :	Sodium Alginate	3g
	Water	300㎖

(B) Second Coating

Seed : Coated granule according to (A)		253g
Coating-Solution:	HPMCP	50g
	Ethanol/Acetone Mixture(1/1, v/v)	700㎖

Example 4

The coated granule according to the present invention was prepared according to the same procedure as Example 1 except that materials as described below were used.

(A) First Coating

Seed : *Lactobacillus acidophilus*: *Lactobacillus bifidus*: *Streptococcus faecalis* = 1:1:1(w/w/w) mixture		250g
Coating-Solution :	Eudragit L30D	300㎖
	Water	150㎖
	Propylene Glycol	9g

Example 5

The coated granule according to the present invention was prepared according to the same procedure as Example 1 except that materials as described below were used.

(A) First Coating

Seed :	*Lactobacillus acidophilus*: *Lactobacillus bifidus*: *Streptococcus faecalis* = 1:1:1(w/w/w) mixture	125g
	Calcium Gluconate	125g
Coating-Solution :	Sodium Alginate	3g
	Water	300㎖

(B) Second Coating

Seed :	Coated granule according to (A)	253g
Coating-Solution:	Zein-DP(Processed from Corn protein extract)	30g
	80% Ethanol	500㎖
	Glycerine	5㎖

Example 6

The coated granule according to the present invention was prepared according to the same procedure as Example 1 except that materials as described below were used.

(A) First Coating

Seed : *Lactobacillus acidophilus*: *Lactobacillus bifidus*: *Streptococcus faecalis* = 1:1:1(w/w/w) mixture		125g
Xylitol		125g
Coating-Solution :	Sodium Alginate	3g
	Water	300mℓ

(B) Second Coating

Seed : Coated granule according to (A)		253g
Coating-Solution:	Chitin	25g
	Water	500mℓ
	Triethyl Citrate	3g
	Acetic Acid (to control the pH value of solution to 2.5 to 3.0)	q.s.

Example 7

The coated granule according to the present invention was prepared according to the same procedure as Example 1 except that materials as described below were used.

(A) First Coating

Seed : *Lactobacillus acidophilus*:

	Lactobacillus bifidus:	
	Streptococcus faecalis	
	= 1:1:1(w/w/w) mixture	125g
	Galacto-oligosaccharide	125g
Coating-Solution :	Sodium Alginate	3g
	Water	300㎖

(B) Second Coating

Seed : Coated granule according to (A)		253g
Coating-Solution:	Carbopol 940(Carbomer® 940)	10g
	Water	500㎖

Example 8

The coated granule according to the present invention was prepared according to the same procedure as Example 1 except that materials as described below were used.

(A) First Coating

Seed :	*Lactobacillus acidophilus*:	
	Lactobacillus bifidus:	
	Streptococcus faecalis	
	= 1:1:1(w/w/w) mixture	125g
	Calcium Gluconate	125g
Coating-Solution :	Sodium Alginate	3g
	Water	300㎖

(B) Second Coating

Seed : Coated granule according to (A)		253g
Coating-Solution:	Soybean Protein	30g
	Water	500㎖
	(phosphate buffer, pH 7.2)	
	Glycerine	5㎖

Example 9

The coated granule according to the present invention was prepared according to the same procedure as Example 1 except that materials as described below were used.

(A) First Coating

Seed : *Lactobacillus acidophilus*:		
Lactobacillus bifidus:		
Streptococcus faecalis		
= 1:1:1(w/w/w) mixture		125g
Mannitol		125g
Coating-Solution :	Sodium Alginate	3g
	Water	300㎖

(B) Second Coating

Seed : Coated granule according to (A)		253g
Coating-Solution:	Xanthan Gum	20g
	Water	500㎖
	Glycerine	5㎖

Experimental Example 1

In order to examine whether the coated granules prepared in Examples 1 to 9 exhibit any changes in artificial gastric juice and intestinal juice [which are prepared according to USP], the following in vitro experiments were conducted. Then, the results thus obtained were compared with those of commercially available products, Dr. Capsule (Binggrae Co.) and *Bifidus* strain original powder-1 (10^8 times) (Cell Biotech. Co.).

First, 10g of each of coated lactic acid bacteria was stirred in 100㎖ of artificial gastric juice for one hour at 50rpm and then the residue was transferred to 100㎖ of artificial intestinal juice. The coated lactic acid bacteria were slowly stirred for 5 hours in artificial intestinal juice and then incubated (cuture medium: Elliker broth; curture condition: anaerobic, 37℃, 72 hours). Then, the disintegration degree of lactic acid bacteria was determined by checking the time when a spongy phase was macroscopically observed. In Table 2, the disintegration data in artificial intestinal juice means the time when 100% of the coated granule is disintegrated and the survival rate was calculated according to the following equation:

$$\text{Survival rate} = \frac{A}{B} \times 100$$

In the above equation,

A represents the number of lactic acid bacteria obtained by stirring for one hour in gastric juice and for 5 hours in intestinal juice and then incubating, and

B represents the number of lactic acid bacteria obtained by stirring

for 5 hours only in intestinal juice and then incubating.

Each of the results represented in Table 2 is an average value of three runs.

Table 2.

Acid-resistance(survival rate) and disintegration data of coated lactic acid bacteria

	Disintegration		Survival Rate(%)	Remarks
	In artificial gastric juice (one hour; pH 1.2)	In artificial intestinal juice (5 hours; pH 6.8)		
Example 1	No change	Within 3 hours	65	
Example 2	No change	Within 2 hours	43	
Example 3	No change	Within 2 hours	55	
Example 4	No change	Within 1 hour	35	
Example 5	No change	Within 3 hours	23	
Example 6	No change	Within 2 hours	31	
Example 7	No change	Within 1 hour	27	
Example 8	No change	Within 2 hours	25	
Example 9	No change	Within 2 hours		
Product A	No change	More than 5 hours	17	Gelatin coating
Product B	No change	Within 1 hour	3	

Note) Product A : Dr. Capsule (Binggre Co., Korea)

Product B : Pasteur VIP (Pasteur Co., Korea)

As can be seen from the results given in the above Table 2, the coated granule of lactic acid bacteria of the present invention exhibits a superior survival rate in artificial gastric juice and further, can be disintegrated rapidly in the intestine, in comparison with the commercially available prior products. Therefore, the coated granule of lactic acid bacteria as prepared according to the present invention is recognized as the optimal form which can regulate the in vivo activity of lactic acid bacteria in the best manner.

WHAT IS CLAIMED IS:

1. An enteric coated granule prepared by coating lactic acid bacteria-containing seed with a water-miscible coating material.

2. The coated granule according to claim 1, wherein lactic acid bacteria is one or more selected from the group consisting of the strains belonging to *Streptococcus* genus, *Lactococcus* genus, *Leuconostoc* genus, *Pediococcus* genus, *Enterococcus* genus, *Lactobacillus* genus and *Bifidobacterium* genus.

3. The coated granule according to claim 1, wherein the water-miscible coating material is one or more selected from the group consisting of sodium alginate, alginic acid, polymethylmethacrylate, wheat protein, soybean protein, methylcellulose, hydroxypropylcellulose, hydroxy-propylmethylcellulose, polyvinylacetate phthalate, guar gum, locust bean gum, xanthan gum, gellan gum and arabic gum.

4. The coated granule according to claim 3, wherein the water-miscible coating material is sodium alginate.

5. The coated granule according to claim 3 or 4, wherein the water-miscible coating material is used in an amount of 1 to 80% by weight with respect to the seed.

6. The coated granule according to claim 1, wherein the seed further contains one or more substances selected from the group consisting of starch, lactose, oligosaccharides, glycoalcohols, calcium gluconate, calcium lactate and gluconic acid.

7. The coated granule according to claim 1, wherein after coating

with a water-miscible coating material the coated granule is further coated with a controlled-release coating material.

8. The coated granule according to claim 7, wherein the controlled-release coating material is one or more selected from the group consisting of corn protein extract and processed materials thereof, sodium alginate, alginic acid, polymethylmethacrylate, shellac, hydroxypropylmethylcellulosephthalate, hydroxypropylmethylcellulose, hydroxypropylmethylcellulose acetate succinate, carboxymethycellulose, hydroxypropylcellulose, celluloseacetatephthalate, polyvinylacetatephthalate, ethylcellulose, methylcellulose, soybean protein, wheat protein, chitin, chitinic acid, agar, carrageenan, pectin, carbopol, guar gum, locust bean gum, xanthan gum, gellan gum and arabic gum.

9. The coated granule according to claim 8, wherein the controlled-release coating material is one or more selected from the group consisting of corn protein extract, hydroxypropylmethylcellulosephthalate and shellac.

10. The coated granule according to claim 8 or 9, wherein the controlled-release coating material is used in an amount of 1 to 95% by weight with respect to the granule first coated with a water-miscible coating material.

11. The coated granule according to claim 7, wherein one or more solvents selected from the group consisting of water, alcohol, acetone, acetonitrile, methylene chloride, ether, hexane, chloroform, 1,4-dioxane, tetrahydrofuran, dimethylsulfoxide, ethyl acetate and methyl acetate are used for coating with a controlled-release coating material.

12. The coated granule according to claim 1 or 7, wherein one or

more plasticizers selected from the group consisting of polyethylene-glycols, myvacet, propyleneglycol, glycerine, triethyl citrate, triacetin, cetyl alcohol and stearyl alcohol are mixed with the coating material.

13. The coated granule according to claim 12, wherein the plasticizer is used in an amount of 1 to 50% by weight with respect to the coating material.

14. The coated granule according to claim 1 or 7, wherein the coating procedure is carried out at the temperature ranging from 20 to 55℃.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/KR 98/00314

A. CLASSIFICATION OF SUBJECT MATTER

IPC[6]: C 12 N 11/02,11/08,11/10,11/12 // (C 12 N 11/02; C 12 R 1:46,1:225)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC[6]: C 12 N 11/02,11/08,11/10,11/12

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

WPI, EPODOC, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	Patent Abstracts of Japan, Vol.12, No.56, 1988, JP 62-201 823 A (FREUNT IND. CO., LTD.) 19 February 1988 (19.02.88).	1-3
A	Database WPI, week 9748, London: Derwent Publications Ltd., AN 97-513517, CN 1 124 773 A (HUNAN COLLEGE TRADITIONAL CHINESE MEDICI), abstract.	1

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

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"O" document referring to an oral disclosure, use, exhibition or other means

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"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
01 February 1999 (01.02.99)	16 February 1999 (16.02.99)
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